



IPEX Syndrome and Gene Therapy Case Study

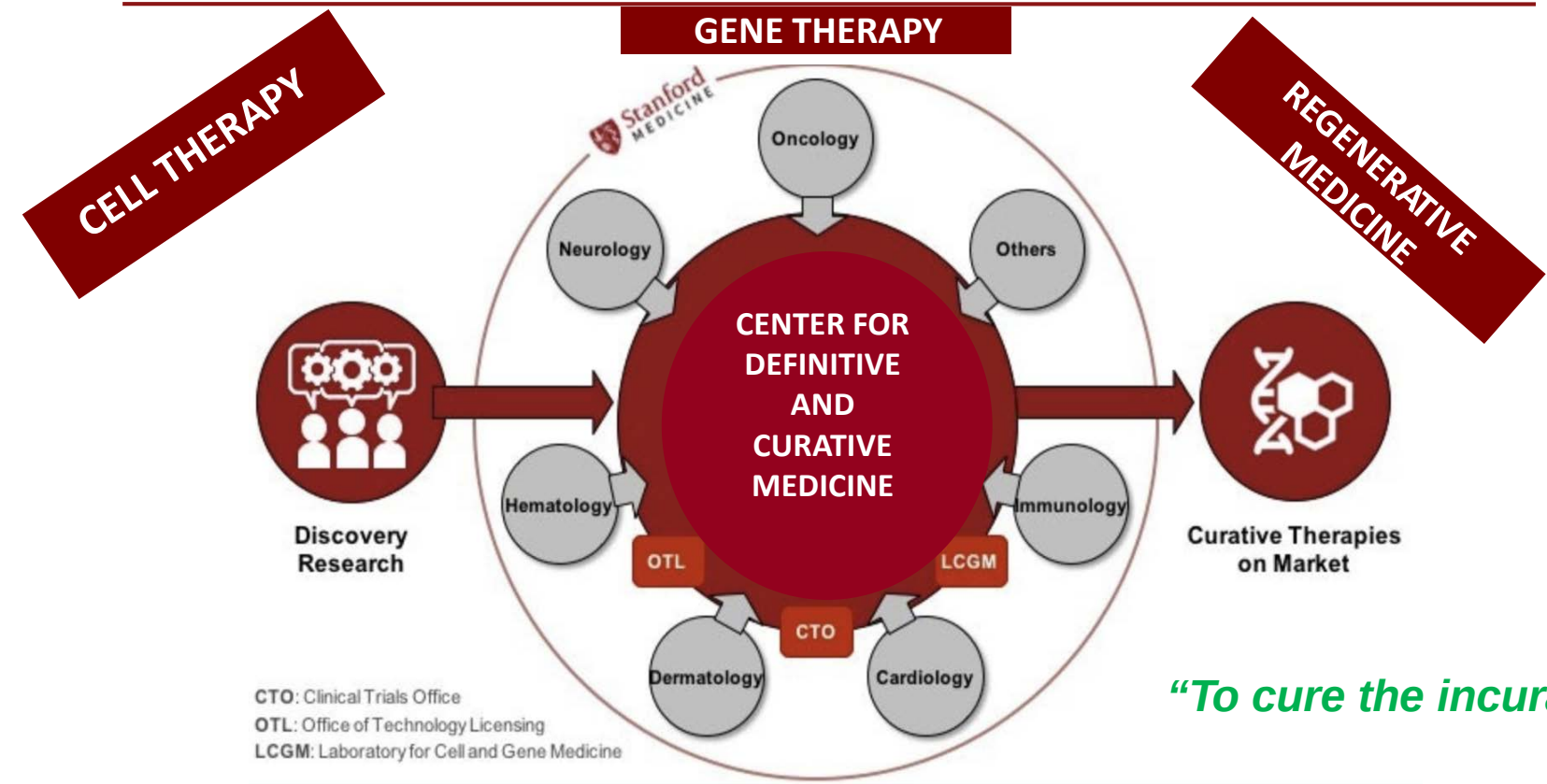
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Immune Dysregulation Polyendocrinopathy Enteropathy X-linked (IPEX) Syndrome

Prototype of Genetic Autoimmune Disease

Genetic disease: FOXP3 gene mutation resulting in regulatory T cell dysfunction

Autoimmune Clinical Manifestations: neonatal diabetes, severe enteropathy and eczema

Severe onset and outcome: fatal in infancy

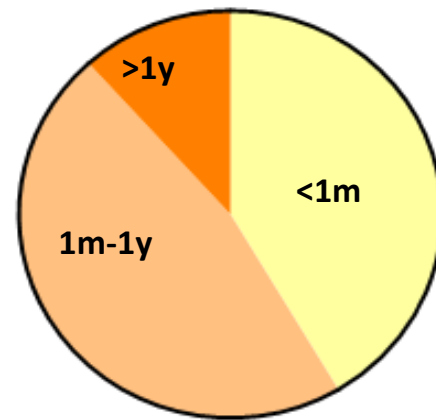
Current treatments: only partial efficacy



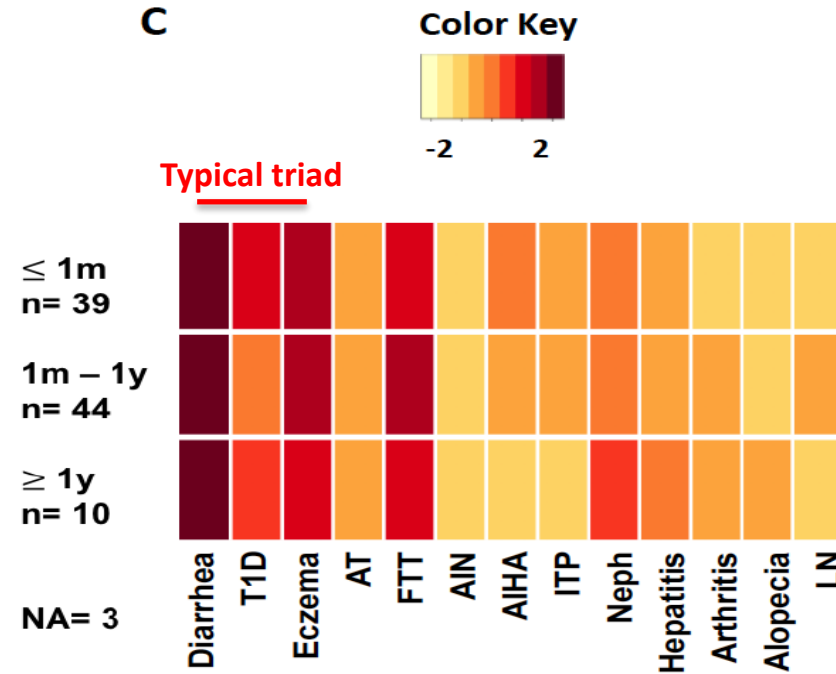
Rare disease with wide distribution-Incidence under estimated

Bacchetta et al, Ann Rev NYAS 2016; Barzaghi et al, JACI 2018

Immune dysregulation Polyendocrinopathy Enteropathy X-linked Syndrome (IPEX): main features at onset



age of onset
N=96 patients



Barzaghi F, Frontiers Immun, 2012
Bacchetta R, NYAS 2016
Barzaghi F, JACI 2018

TWO BROTHERS WITH THE SAME FOXP3 MUTATIONS, BUT DIFFERENT CHANCES TO BE CURED

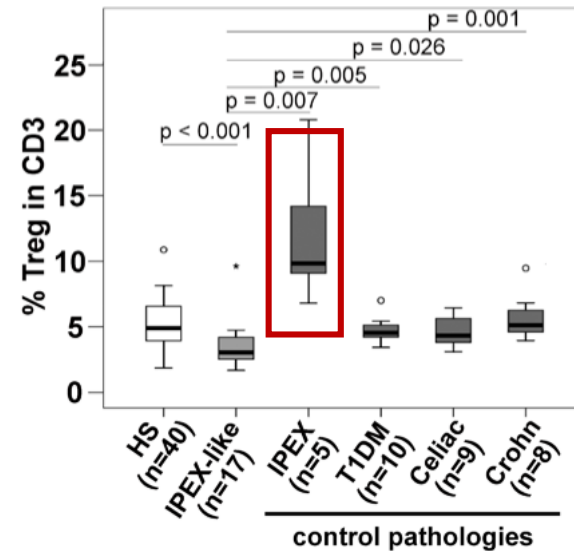
- **Onset at few weeks of life**, with severe enteropathy, severe weight loss and hepatitis
- **Genetic diagnosis of FOXP3 mutation** at about 4mo, *after already 3 mo in hospitals*
- **Immune suppression over the course of 18 months**: methylprednisolone, tacrolimus, sirolimus, ATG, ATG, tocilizumab, bortezomib, and campath. Overall poor response.
- **$\alpha\beta$ T-cell depleted haploidentical HSCT** from the father **at 18 months**
- **Today**: 3 months post HSCT: alive, 100% donor chimerism, no IS, *he is carrying over gut and liver problems.*

- **Onset at few weeks of life**, admitted to Stanford ER with severe dehydration and weight loss.
- **Immunologic diagnosis within the first month and genetic diagnosis** of FOXP3 mutation confirmed 2 weeks later.
- Presence of autoAb but no clinical manifestations yet.
- **Immune suppression** : methylprednisolone, and sirolimus with good response.
- **$\alpha\beta$ T-cell depleted haploidentical HSCT** from the father **at 4 months of age.**
- **Today** 1 month post HSCT: alive, engrafted, no IS, no autoimmune clinical manifestations.

.....need for early diagnosis and curative therapy available to every patients

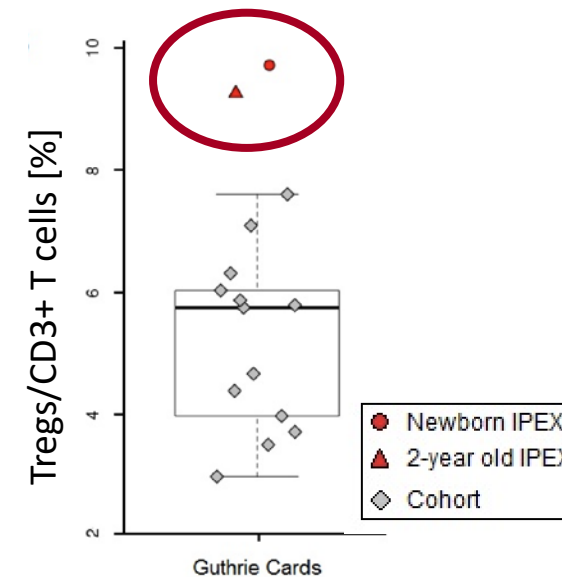
IPEX syndrome can be identified at birth: facilitate approval of tests that can be broadly used to identify the disease

- Combined epigenetic quantification of TSDR for regulatory T cell and TLSDR for CD3+ T cells allows discrimination between IPEX and IPEX-like patients.



Barzaghi et al., J Autoim 2012

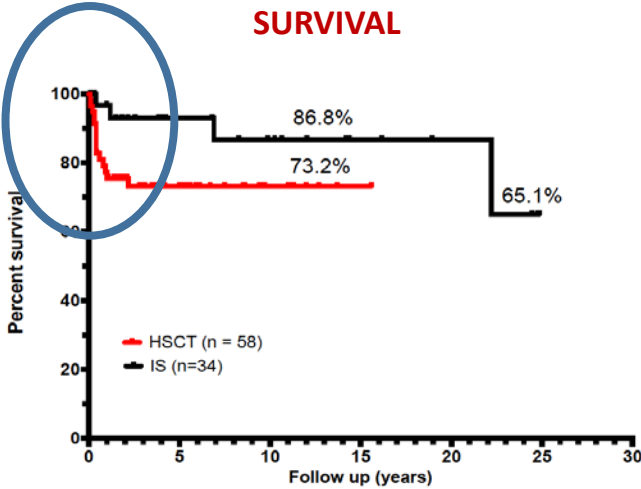
- Potential for newborn screening



Baron U, Sci Transl Med 2018

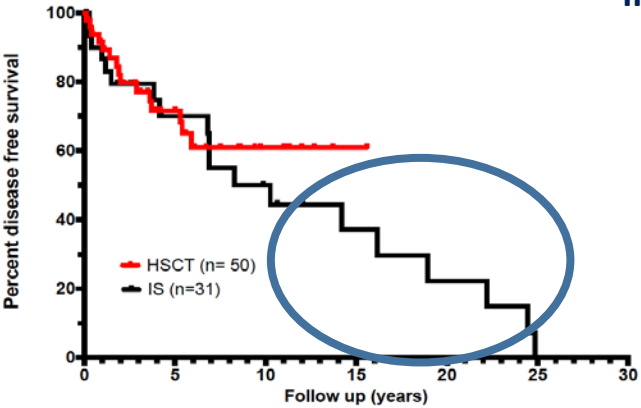
What can be plan to improve current treatment outcome

A. Intervene at onset to bridge to transplant or instead of the transplant



	Survival probability (%)				
Time	6m	1y	5y	10y	15y
HSCT	82.8	75.5	73.2	73.2	73.2
IS	96.7	96.7	92.9	86.8	86.8

DISEASE FREE SURVIVAL



	Survival probability (%)				
Time	6m	1y	5y	10y	15y
HSCT	93.9	89.3	71.6	61.0	61.0
IS	90.0	83.1	65.1	50.1	37.1

B. Intervene in chronically ill patients to reduce the continue use of immune suppressive drugs

Two possible approaches of cell/gene therapy for IPEX syndrome

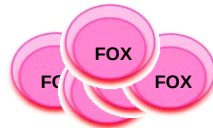
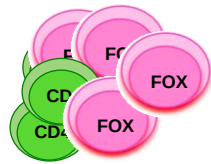
Cell therapy with
LV-engineered T cells
to restore tolerance

wtFOXP3 over-expression in
conventional CD4⁺ T cells

LV-FOXP3/NGFR



in vitro expansion and
selection of transduced cells



Sato Y, Postdoct

Passerini L et al. STMed 2013

Re-infusion of purified
autologous CD4^{LVFOXP3} T cells
with normal regulatory function

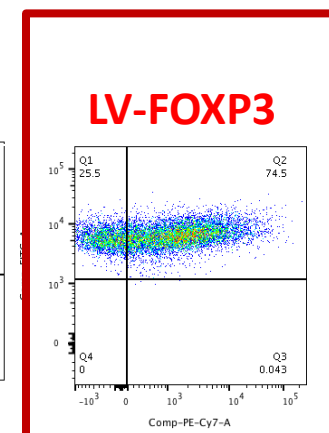
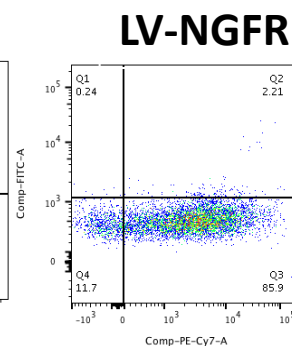
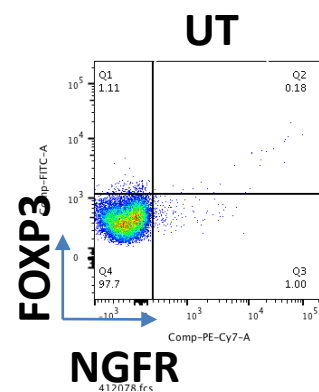
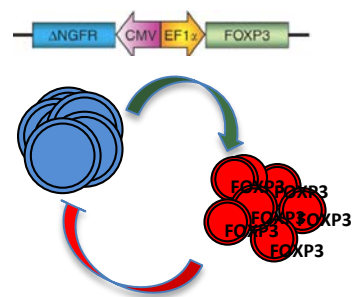
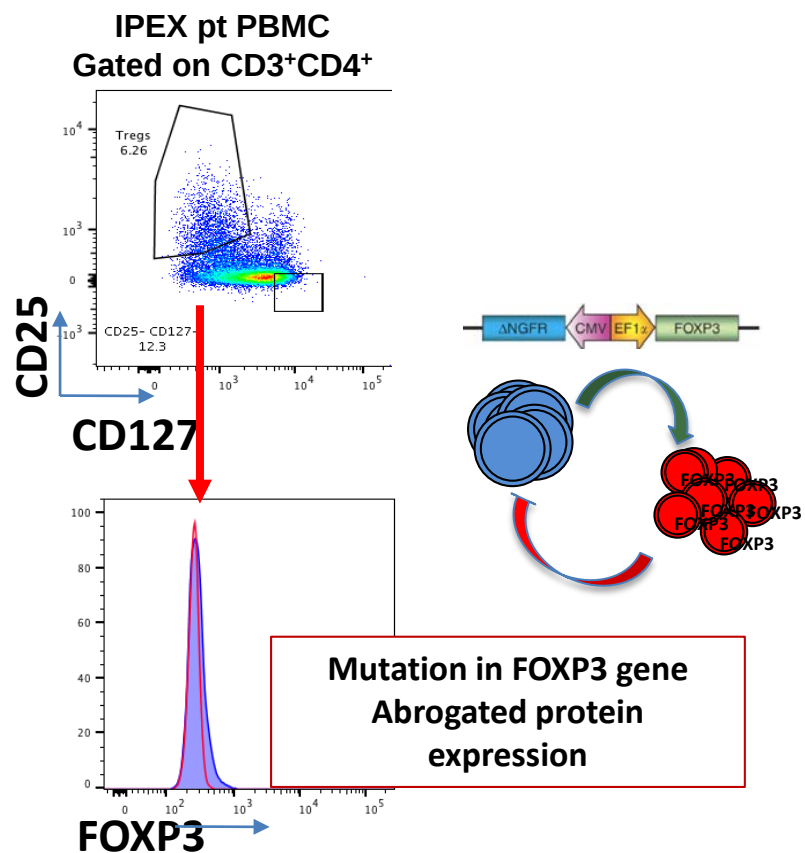
Gene therapy with
CRISPR/Cas9 gene corrected
autologous HSPCs to preserve
endogenous regulation of FOXP3
gene expression



• Ongoing studies on best
strategy

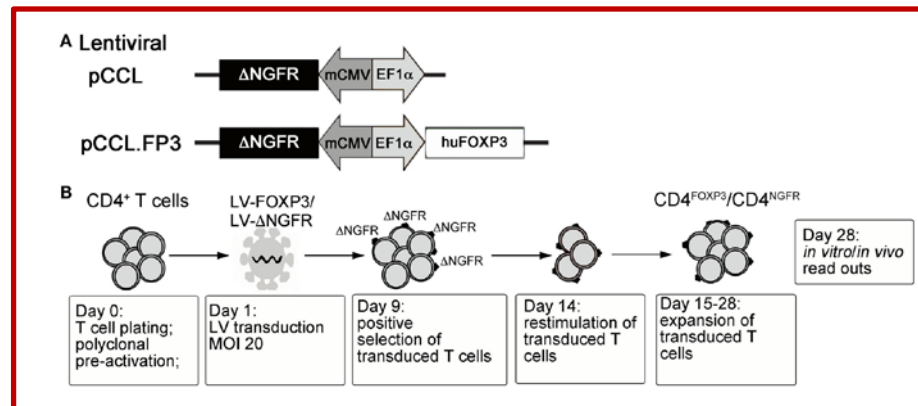
Bacchetta Lab, Stanford

CD4^{LV-FOXP3} “CONVERTED” Treg CAN BE OBTAINED FROM FOXP3 null T CELLS

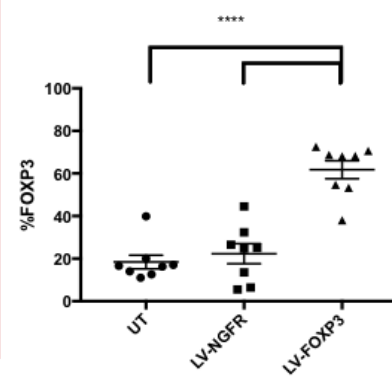


**FOXP3 gene protein expression
Restored after LV-FOXP3 “conversion”**

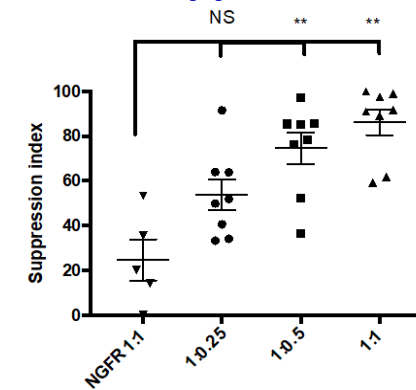
DEVELOPING CD4^{LVFOXP3} “CONVERTED” TREG CELLS FOR CLINICAL USE



FOXP3 expression



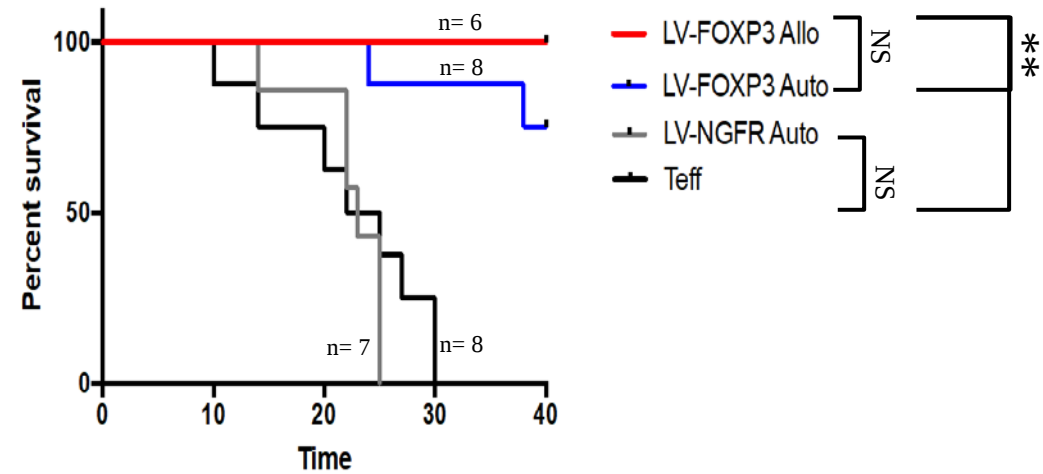
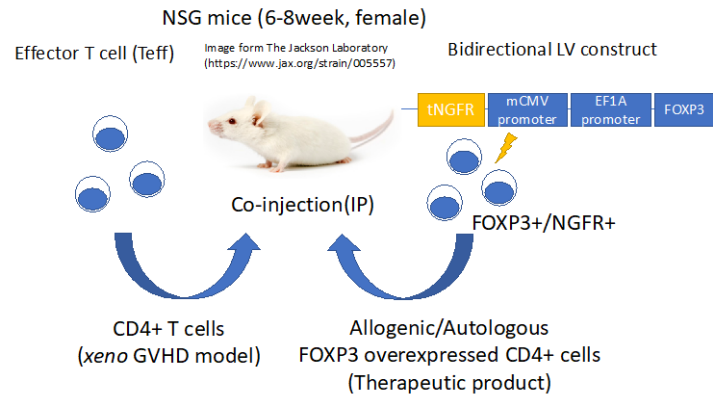
Suppression



.....process is consistent and reproducible among different subjects

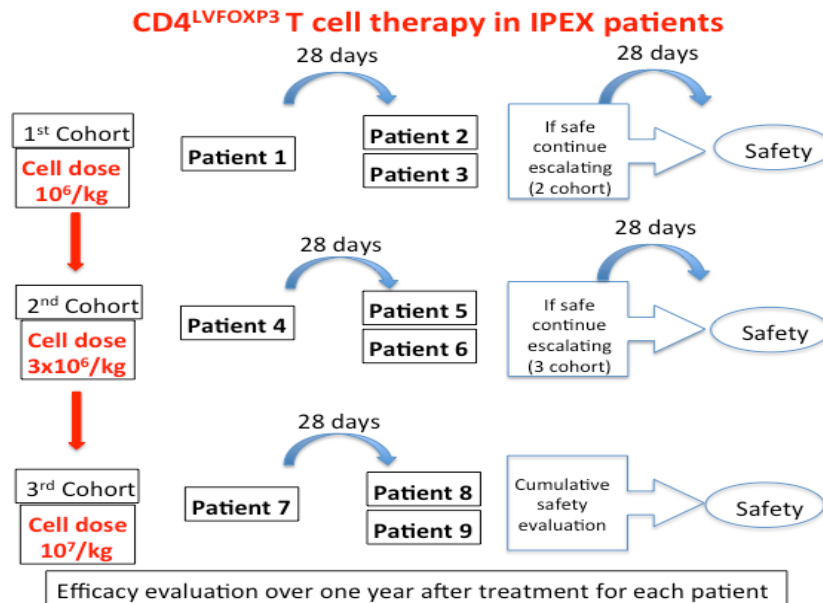
DEVELOPING $CD4^{LVFOX P3}$ “CONVERTED” TREG CELLS FOR CLINICAL USE

Methods: *in vivo* (xeno GVHD model)



Safety, efficacy, survival tested in hu-mice models

PHASE I CLINICAL PROTOCOL FOR THE CLINICAL USE OF CD4^{LV}FOXP3 T CELLS IN IPEX SYNDROME



Primary Objective:

- Asses safety
- Dose ranges and feasibility
- PK and PD of the cell product

Outstanding Questions:

- Patient inclusion criteria?
- Disease status
- Age
- Response to prior therapy

KEY QUESTIONS TO EASE THE CLINICAL APPLICABILITY OF PHASE I TRIALS IN IPEX AND SIMILAR BLOOD RARE DISEASES

- **CRITERIA FOR PATIENTS ELIGIBILITY FOR RARE DISEASES**
Usual criteria for phase 1 study include the presence of clinical manifestation and failed previous therapy: based on clinical history, tools for early diagnosis and disease measurements, *can we aim to treat even before the symptoms arise?*
- **AGE CONSTRAIN FOR PHASE I TRIAL:**
many of these disease have very early onset, patients can deteriorate very quickly: *can we intervene at early pediatric age?*
- **DISEASE SPECIFIC CRITERIA** for evaluating safety and efficacy of a cell/gene product based on preclinical model: what are the expectations from the available preclinical models?
- **Need to establish a committee to look at the issues related to clinical research in rare disease.**

Stanford University

Pediatric Stem Cell Transplantation and Regenerative Medicine Team, Stanford University

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Division Chief
CO-directore CDCM

Translational Medicine
Immune Tolerance
Gene Therapy



Matthew Porteus, MD, PhD
CO-directore CDCM

Genome Editing
Hemoglobinopathies
PIDs



Robertson Parkman, MD

Immune Reconstitution
SCT for PIDs



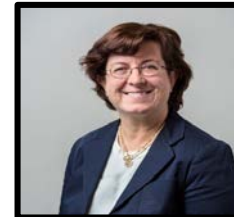
Ken Weinberg, MD

Immune Reconstitution
Viral Specific Immunity
HSC Biology



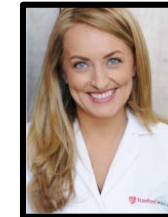
Rajni Agarwal-Hashmi, MD

Experimental SCT
CAR-T Therapies



Rosa Bacchetta, MD

Genetic Autoimmune Diseases
Tolerance Induction
Genome Editing



Agnieszka Czechowicz, MD, PhD

BMF/Fanconi Anemia
mAb Conditioning



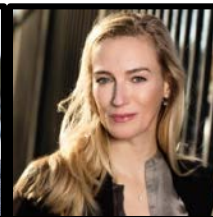
Ami Shah, MD

Quality Control
SCT and Gene Therapy
for Metabolic Diseases



Sandeep Soni, MD

Hemoglobinopathies
Gene Therapy



Katja Weinacht, MD, PhD

Pathophysiology of PID's
Autoimmune Anemias
HSC Metabolism



Alice Bertaina, MD, PhD

Haplo- Transplantation
Experimental SCT



Melissa Mavers, MD, PhD

GvHD Pathophysiology
GvHD Treatment



Julia Chu, MD

Immune Reconstitution
Gene Therapy for PIDs